



Supplemental Supplemental Figure 1. Expression of GPCRs in TCGA invasive breast cancer dataset. A, $G_{i/o}$ -coupled GPCRs; B, G_s -coupled GPCRs; C, $G_{q/11}$ -coupled GPCRs; D, $G_{12/13}$ -coupled GPCRs; E, GPCRs with unknown G-protein linkage. The expression levels (high, low and no change) of GPCR mRNA relative to diploid samples are organized based on the molecular subtypes of breast cancer shown on the top.

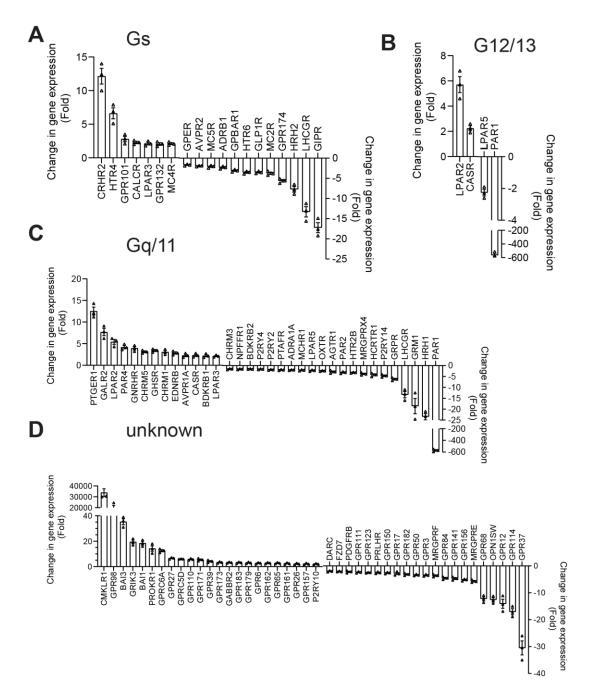
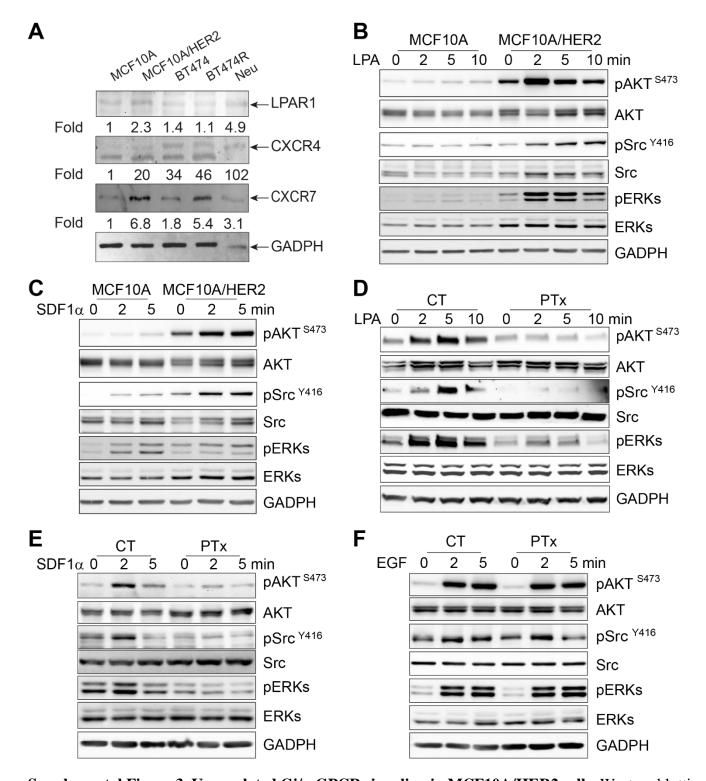
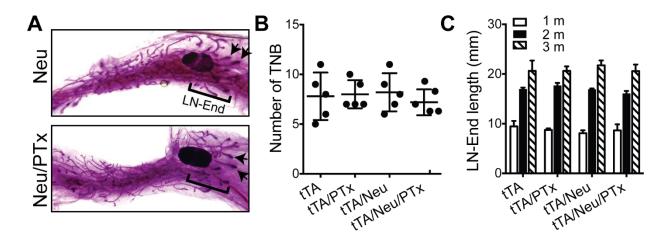


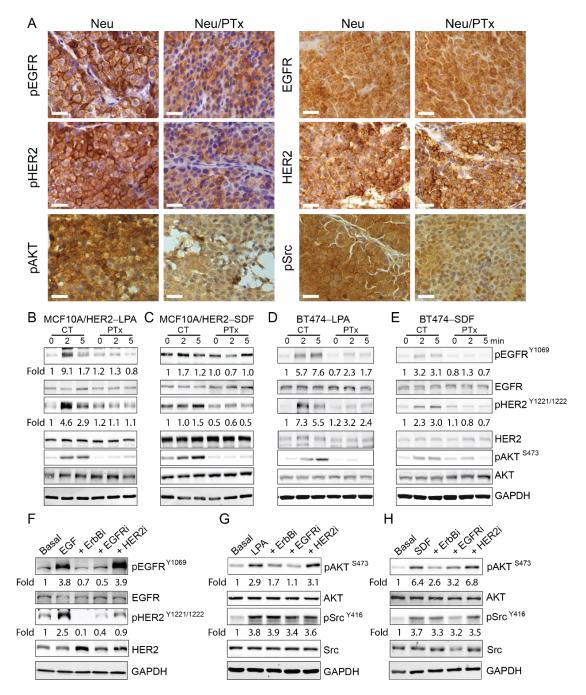
Figure 2. Altered GPCR expression in Neu cells. A, G_s -coupled GPCRs; B, $G_{q/11}$ -coupled GPCRs; C, G_s -coupled GPCRs; D, GPCRs with unknown G-protein linkage that show more than 2-fold change of expression in Neu cells as compared to control cells. The data are expressed as mean \pm SEM, n=3.



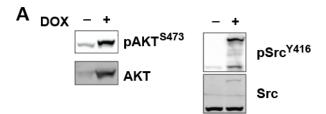
Supplemental Figure 3. Upregulated Gi/o-GPCR signaling in MCF10A/HER2 cells. Western blotting showing A, the expresson of LPAR1, CXCR4 and CXCR7 in the indicated cells. The levels of LPAR1, CXCR4 and CXCR7 in each cell line were quantified as the ratio of the protein to GADPH and expressed as the fold increase over MCF10A, which is indicated underneath the images; B, the response of MCF10A and MCF10A/HER2 to LPA (B)- and SDF1 α (C)-stimulated AKT^{S473} and Src^{Y416} phosphorylation; D-F, the effect of PTx treatment on LPA (D)-, SDF1 α (E)- and EGF (F)-stimulated AKT^{S473} and Src^{Y416} phosphorylation in MCF10A/HER2 cells. The images are representative of at least three independent experiments and were assempled from multiple blots run with samples from the same experiments.

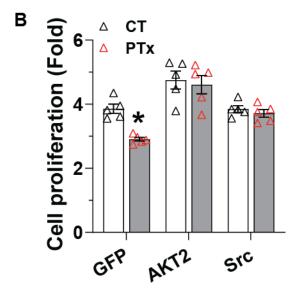


Supplemental Figure 4. PTx expression does not affect mammary gland development. A, Whole-mount *in situ* staining showing the terminal end buds (TNBs, indicated by arrows) and the length of ductal distance, measured from the lymph node (LN) to the end of TNBs (LN-End), in the mammary glands from 1-month old Neu and Neu/PTx mice. B-C, quantitative data showing the number of TNBs (B) and the length of LN-End (C) in mammary glands from transgenic mice at different ages. M: month.



Supplemental Figure 5. Cross-talk between $G_{i/o}$ -GPCRs and EGFR/HER2. A. IHC staining showing phospho-EGFR Y1069 (pEGFR), total EGFR, phospho-HER2 $^{Y1221/1222}$ (pHER2), total HER2, phospho-AKT S473 (pAKT), and phospho-Src Y416 (pSrc), in Neu and Neu/PTx tumors. Scale bar=10 μm. The images are represtative of 6 to 9 tumors. B-E, Western blotting showing phosphorylation of EGFR, HER2, and AKT in MCF10A/HER2 (B-C) and BT474 (D-E) cells stimulated with 10 μM LPA (B) and 50 nM SDF1α (C) and treated with vehicle (CT) or PTx (200ng/ml). D-F, the effect of 1 μM of the pan-ErbB-, EGFR-, and HER2-selective inhibitors on the activation of EGFR and HER2 by EGF in Neu cells (D) and the phosphorylation of AKT S473 and Src Y416 by LPA (E) and SDF1 α (F) stimulation in MCF10A/HER2 cells. The phosphorylation of EGFR Y1068 , HER2 $^{Y1221/1222}$, AKT S473 , and Src Y416 was quantified as the ratio of the phosphorylated to total protein and expressed as the fold increase over basal, which is indicated underneath the images. All of the blotting images are representative of at least three independent experiments and assembled from multiple blots run with the samples from the same experiments.





Supplemental Figure 6. $G_{i/o}$ -GPCRs signal through AKT and Src pathways to regulate Neu cell growth. A, Western blotting showing the expression of myristoylated AKT2 and GFP-tagged Src/Y527F in Neu cells induced by doxycylcine (DOX, 0.5 µg/ml for AKT2 and 0.1 µg/ml for Src) for three days. The images were assembled from multiple blots run with the samples from the same experiment. B, the effect of induced myristoylated AKT2 and GFP-tagged Src/Y527F expression on PTx-mediated inhibition of Neu cell growth. Neu cells expressing the indicated constucts were treated with doxycyline in the presence of vehicle (CT) or PTx (200ng/ml) for 5 days. Two-tail unpaired Student's t test, *p<0.05, n=5.